Phytopharmacology



Neuropharmacological effects of triterpenoids

Parmar SK^{1,*}, Sharma TP¹, Airao VB¹, Bhatt R¹, Aghara R¹, Chavda S¹, Rabadiya SO¹, Gangwal AP²

¹Division of Pharmacology, Department of Pharmaceutical Sciences, Saurashtra University, Rajkot 360 005, Gujarat, India.

²Smriti College of Pharmaceutical Education, Indore 452 010, Madhya Pradesh, India.

Corresponding author: parmarsachin@rediffmail.com; Tel: +919898002327; Tele-Fax: +91 2812585083

Received: 10 November 2012, Revised: 23 January 2013, Accepted: 28 January 2013

Abstract

Triterpenes comprise one of the most interesting groups of natural products due to their diverse pharmacological activities. Triterpenes are ubiquitously present in variety of ethnomedicinal plants. The term 'triterpene' represents naturally occuring terpenes, whereas the broader expression 'triterpenoid' includes secondary metabolites. It has been estimated that 80 distinct types of both the structure and the chemical characteristics of triterpenes have been identified till today. Many such compounds can either be used directly as active compounds or modified to increase their selectivity and potency. The present article provides updates on wide range of biological activities of tetracyclic triterpenes and pentacyclic triterpenoids such as immunomodulatory, anticancer, anti-inflammatory, anti-anxiety, antidepressant, memory enhancer, antinociceptive, neuroprotective and other CNS actions. Several structural groups of triterpenes have demonstrated specificity against transcriptional factors which can be promising candidates for treating inflammation, cancer, and immune diseases.

Keywords: Ethnomedicine; Molecular signaling; Neuropharmacology; Pentacyclic triterpenoids; Tetracyclic triterpenes

Introduction

Triterpenoids are widely distributed in the plant kingdom. They are produced in plant as secondary metabolites and have varied biological activities (Hanson, 2003). The terms triterpenes and triterpenoids are often used to describe the same C_{30} -terpene compound. However, they need to be differentiated based upon their occurrence, biosynthesis and biotransformation products. The term 'triterpene' is used to describe naturally occurring terpenes whereas; the broader expression 'triterpenoid' includes natural degradation products (Eggersdofer, 2005). Triterpenes are originally synthesized by plants as metabolites, and are abundantly present in the plant kingdom in the form of free acids or aglycones (Chappell, 1995; McGarvey Croteau, 1997). Till today, at least 80 distinct types of both the structure and the chemical characteristics of triterpenes have been shown. It is well-recognized that triterpenes have long been used as flavors, pigments, polymers, fibers, glues, and waxes. In many Asian countries, herbal products containing triterpenes are widely prescribed to prevent or treat a variety of diseases by the traditional healers (Wagner and Elmadfa, 2003; Xu et al., 2004).

Classification of triterpenes and triterpenoids

Triterpenoids are structurally diverse group of natural products that contain about 30 carbon atoms. As shown in Table 1, the triterpenoids are classified into two main groups; tetracyclic and the pentacyclic triterpenoids. Tetracyclic triterpenes such as oleandrin, euphol and cucubitacins are methylated steroids (Figure 1). The group of pentacyclic triterpenoids is by far the largest and friedelane, lupane, ursane, oleane, serratane, and taraxastane are six main groups of this category (Gunatilaka, 1986; Xu et al., 2004). Ursanes and oleananes such as oleanolic acid, ursolic acid, maslinic acid, uvaol, and erythrodiol (Figure 1) are the major triterpene skeletons present in higher plants including commonly consumed plant foods (Abe et al., 1993; Connolly and Hill, 2001). Other groups of triterpenes occur widely in edible or inedible plants (Yin M., 2012).

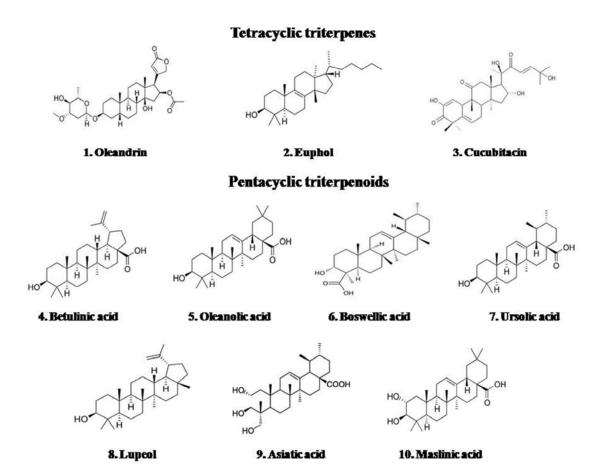


Figure 1. Chemical structures of tetracyclic triterpenes and pentacyclic triterpenoids.

Phytopharmacology 2013, 4(2), 354-372

Triterpenoid Family	Triterpene	R ₁	R ₂
	Ursolic acid	СООН	
Ursane	Uvaol	CH ₂ OH	
	α-amyrin	CH ₃	
	Erythrodiol	CH ₂ OH	Н
Oleanana	β-amyrin	CH ₃	Н
Oleanane	Oleanolic acid	СООН	Н
	Maslinic acid	СООН	OH
	Betulin	CH ₂ OH	
Lupane	Lupeol	CH ₃	
	Betulinic acid	СООН	

Table 1. Classification of triterpenoids and their derivatives.

Occurrence in plant kingdom

Although terpenes are widely distributed in the plant kingdom, most of the bioactive terpenes have been found in higher plants. Mono- and sesquiterpenes are chiefly present in plants possessing volatile oil whereas, higher terpenes, such as triterpenes are chiefly discovered in balsams and resins (Gildemeister and Hoffmann, 1960; Sandermann, 1960). They are also present in prokaryotic as well as eukaryotic organisms. An excellent review on bacterial triterpenoids with specific focus on triterpenoid occurrence and their functions in bacteria has been described by Tylor (1984). The triterpenes content of different plants varies depending on parameters such as species, season and soil. Table 2 depicts widespread occurrence of triterpenes and triterpenoids in plants.

Chemical compound	Botanical name	Family
Tetracyclic triterpenoid		
Cucurbitacin	Bryonia alba	Curcutibaceae
Ganoderic acid	Ganoderma lucidum	Ganodermataceae
Oleandrin	Nerium oleander	Apocynaceae
Pentacyclic triterpenoid		
Amyrin	Diospyros kaki	Ebenaceae
Asiatic acid	Centella asiatica	Mackinlayaceae
Avicin	Acacia victoriae	Fabaceae
Betulinic acid	Ziziphus mauritiana	Rahmnaceae
	Anemone raddeana	Ranunculaceae
	Lycopodium cernuum	Lycopodiaceae
	Syzygium claviflorum	Myrtaceae
Boswellic acid	Boswellia serrata	Burseraceae
	Boswellia carteri	Burseraceae
Lupeol	Mangifera indica	Anacardiaceae
	Crataeva nurvala	Capparidaceae

Table 2. Occurrence of triterpenes and triterpenoids in the plant kingdom.

Madecassic acid	Centella asiatica	Mackinlayaceae
Momordin	Kochia scoparia	Amaranthaceae
Oleanolic acid	Arctostaphyllos uva-ursi	Ericaceae
	Calluna vulgaris	Ericaceae
	Crataeva nurvala	Capparidaceae
	Ganoderma lucidum	Ganodermataceae
	Sambucus chinensis	Adoxaceae
	Solanum incanum	Salanaceae
Platycodon D	Platycodon grandiflorum	Campunulaceae
Pristimerin	Maytenus ilicifolia	Celastraceae
	Celastrus hypoleucus	Celastraceae
	Tripterygium wilfordii	Celastraceae
Ursolic acid	Ocimum sanctum L.	Lamiaceae
	Thymus vulgaris L.	Lamiaceae
	Lavandula augustifolia	Lamiaceae
	Nepeta sibthorpii	Lamiaceae
	Mentha piperita L.	Lamiaceae

Biosynthesis

Plants biosynthesize diverse group of triterpenoids. As described in Figure 2, the terpenes biosynthesis can be divided into four distinct stages. Initial or the first stage involves the formation of isopentenyl diphosphate that is basic building block for isoprenoids (Gershenzon and Kreis, 1999). Secondly, these units associate to form the $(C_5H_8)_n$ isoprenoid backbone of the terpene families. Thirdly, there is generation of the carbon skeletons as a result of the cyclization of these units. Finally, various chemical reactions such as hydroxylations and oxidations lead to the formation of individual terpenoids. Most of the triterpenes are derived from squalene, which is synthesized from the reductive coupling of two molecules of farnesyl pyrophosphate by the enzyme squalene synthase. The enzyme squalene epoxidase then oxidize squalene to generate 2, 3-oxidosqualene. Furthermore, these oxidized squalene moiety is cyclized by oxidosqualene cyclases (OSCs) to form intermediate cations. These cations then undergo structural changes by various enzymes to produce triterpene alcohols or aldehydes including α - and β -amyrin and lupeol (Haralampidis et al., 2002; Phillips et al., 2006).

It is well-established that different plants possess genomic machineries for multiple OSC enzymes that facilitate triterpenoid biosynthesis. These OSCs confers the structural diversity to the triterpenoids by their unique role on cyclization of 2,3-oxidosqualene (Mangas et al., 2006). Cyclization of 2,3-oxidosqualene through a protosteryl cation intermediate generates lanosterol and cycloartenol which are structural precursors for all the steroids, whereas cyclization through a baccharenyl, dammarenyl and lupenyl cation intermediates produces lupeol and α/β -amyrin (Jenner et al., 2005). The cyclization of 2,3-oxidosqualene by α/β -amyrin synthase enzymes results into the formation of dammarenyl cation which undergo further ring expansion and a few rearrangements before deprotonation to α -amyrin and β -amyrin, respectively.

Following cyclization, further diversity in structure is conferred by modification of the products by oxidation, hydroxylation, glycosylation and other substitutions mediated by cytochrome P450-dependent monooxygenases, glycosyl transferases and other enzymes. The enzymes employed for these chemical elaborations of triterpenes and triterpenoids have not been well documented.

Phytopharmacology 2013, 4(2), 354-372

Parmar et al.

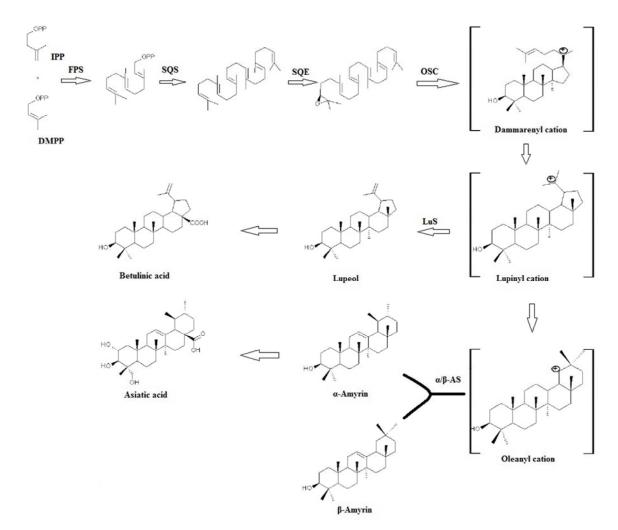


Figure 2. A simplified scheme of triterpenoid biosynthesis. Isomerization of isopentyl diphosphate (IPP) and dimethylallyl diphosphate (DMAPP) by farnesyl diphosphate synthase (FPS) leads to farnesyl diphosphate (FPP), which is then converted to squalene by the enzyme squalene synthase (SQS). Subsequently, squalene epoxidase (SQE) oxidises squalene to 2,3-oxidosqualene. This is followed by enzymatic ciclyzation of 2,3-oxidosqualene by oxidosqualene cyclase (OSC) to produce intermediate dammarenyl cation. Further, the ring expansion of dammarenyl cation produce lupinyl or oleanyl cations. Finally, enzymes α/β -amyrin synthases (α/β -AS) and lupeol synthase converts oleanyl and lupinyl cations to form the α/β -amyrin and lupeol, respectively.

Effects of triterpenoids on molecular and cellular signaling

Biological activities

A wide range of biological activities of triterpenoids have been reported. Some of these activities along with molecular signalling of important tetracyclic and pentacyclic triterpenoids are discussed here. Ursolic acid, a pentacyclic triterpenoid, has been shown to possess immunomodulatory (Jang et al., 2009; Raphael and Kuttan, 2003), antioxidative (Ramachandran and Prasad, 2008), anti-HIV (Lee et al., 2008a), bone anabolic activities (Lee et al., 2008b), hypolipidemic (Min et al., 2008; Somova et al., 2003), antibacterial (Fonranay et al., 2008), anti-mutagenic (Resende et al., 2006), antitumor (Hsu et al., 2004; Ma et al., 2005), antidysrhythmic (Somova et al., 2004), and hepatoprotective (Saraswat et al., 2000). Apop-

tosis inducing activity of betulinic acid have been studied thoroughly in neuroblastoma and glioblastoma cells, which is believed to be mediated by the activation of the mitochondrial pathway (Fulda et al., 1997; Jeremias et al., 2004; Tan et al., 2003).

Table 3. Molecular targets of tetracyclic triterpenes and pentacyclic triterpenoids for antitumor and anti-inflam-
matory actions.

Terpenoids	Targets	References
Tetracyclic triterpenes		
Cucurbitacin	Cyclin B1, cyclin D1, Mcl-1, cdc25C, STAT3, p53	Chan et al., 2010; Lui et al., 2009
Ganoderic acid	NF-KB, AP-1, NFATc1, cdk4, uPA, MMP2, MMP9	Chen et al., 2008; Jiang et al., 2008
Oleandrin	NF-κB, AP-1, Fas, ERK, FGF-1	Afaq et al., 2004; Manna et al., 2000
Pentacyclic triterpenoid	1	
Amyrin	NF-κB, IL-1β, COX-2, CREB, ERK, PKC, P38 MAPK	Medeiros et al., 2007; Vitor et al., 2009
Asiatic acid	NF-κB, caspases-2, -3, -8 and -9, PARP, Bcl-2	Tang et al., 2009; Park et al., 2007
Avicin	NF-κB, Fas, STAT3, caspase-8, Bcl-2, Bcl-xL	Haridas et al., 2009; Zhang et al., 2008
Betulinic acid	NF-κB, STAT3, Bax, Bcl-2, Bcl-xL, FAK	Chen et al., 2008; Chintharlapalli et al., 2007
Boswellic acid	NF-κB, STAT3, AR, p21, DR5, caspase-3 and -8	Kunnumakkara et al., 2009; Syrovets et al., 2005
Celastrol	NF-κB, IAP1, IAP2, Bcl-2, Bcl-xL, c-FLIP, COX-2, survivin, cyclin D1, MMP9, VEGF, iNOS, Hsp90, cdc37, VEGFR	Jung et al., 2007; Kim et al., 2009
Escin	NF-κB, STAT3, JAK2, cyclin D1, Bcl-2, Bcl-xL, survivin, Mcl-1, VEGF, COX-2, MMP9	Harikumar et al., 2010; Tan et al., 2010
Lupeol	NF-κB, cFLIP, survivin, Bax, caspase-3, caspase-9	Murtaza et al., 2009; Lee et al., 2007
Madecassic acid	iNOS, COX-2, TNF-α, IL-1, IL-6	Won et al., 2010
Momordin	NF-κB, AP-1, Bcl-2, Bax, caspase-3, PARP	Hwang., 2005; Kim et al., 2002
Oleanolic acid	NF-κB, mTOR, caspases-3, -8, and -9, ICAM-1, VEGF, PARP, Akt	Chu et al., 2010; Deeb et al., 2008
Pristimerin	NF-κB, PARP-1, JNK, Bax, p27, Bcl-2, Bcl-xL	Tiedemann et al., 2009; Wu et al., 2005
Ursolic acid	NF-κB, STAT3, Bcl-2, Bax, ICAM-1, p53, PKC	Manu et al., 2008; Shishodia et al., 2003

AIF, apoptosis inducing factor; AMPK, 5' AMP-activated protein kinase; AP-1, activator protein-1; Apaf1, apoptotic protease activating factor 1; AR, androgen receptor; Bax, BCL2-associated X protein; Bfl-1/A1, BCL2-related protein A1; cdc, cell division cycle; cdk, cyclin-dependent kinase; cFLIP, cellular FLICE inhibitory protein; COX-2, cyclooxygenase-2; CREB, cAMP response element binding protein; DR, death receptor; EGFR, epidermal growth factor receptor; Egr-1, earyl growth response factor-1; ERK, extracellular signal-regulated kinase; FAK, focal adhesion kinase; FasL, Fas-ligand; FGF-1, fibroblast growth factor-1; GSK3 β , glycogen synthase kinase-3 β ; HMG-CoA, 3-hydroxy-3-methylglutaryl-coenzyme A; Hsp, heat shock protein; IAP, inhibitor of apoptosis protein; ICAM-1, intercellular adhesion molecule-1; IFN- γ , interferon- γ ; IL-1, interleukin-1; iNOS, inducible nitric oxide synthase; JNK, c-Jun N-terminal kinase; MAPK, mitogen-activated protein kinase; Mcl-1, myeloid cell leukemia-1; MCP, monocyte chemotactic protein; MEK, MAPK/ERK kinase, MIP-2, macrophage-inflammatory protein-2; MMP, matrix metalloproteinase; mTOR, mammalian target of rapamycin; NF-AT, nuclear factor of activated T-cells; NF- κ B, nuclear factor-kappa B; PARP, poly (ADP-ribose) polymerase; PI3K, phosphoinositide-3 kinase; PKC, protein kinase C; PPAR, peroxisome proliferator-activated receptor; Sp1, specificity protein 1; STAT3, signal transducer and activator of transcription 3; TF, tissue factor; TLR2, Toll-like receptor-2; TNF- α , tumor necrosis factor- α ; TRAF1, TNF receptor-associated factor-1; uPA, urokinase-type plasminogen activator; VCAM-1, vascular cell adhesion molecule-1; VEGF, vascular endothelial growth factor; VEGFR, VEGF receptor.

Table 3 depicts different molecular targets for tetracyclic and pentacyclic triterpenoids. It can be observed that triterpenoids act on wide range of chemokines and apo-ptotic factors and play major role in tumor suppression, inflammatory response, and immune response.

Behavioral and psychopharmacological effects

Biological effects of pentacyclic triterpenoids

Memory and dementia

Nasira et al. (2011) have investigated the effects of acute administration of asiatic acid isolated from *Centella asiatica* on memory and learning using active and passive avoidance models in experimental animals and concluded that administration of asiatic acid facilitated passive avoidance on memory and learning but had no effect on active avoidance on memory. In another study, ursolic acid has been reported to possess antioxidant activity. Furthermore, the protective effect of ursolic acid against the D-galactose-induced neurotoxicity and learning and memory impairment has been demonstrated. The study also postulated molecular mechanism and concluded that the cerebroprotective action of ursolic acid against D-galactose-induced neurotoxicity might be caused, at least in part, by the restoration of antio-xidant enzyme levels such as super oxide dismutase, catalase, glutathione peroxidase and glutathione reductase with a marked reduction in lipid peroxidation. Additionally, ursolic acid might have increased the level of growth-associated protein GAP43 in the brain of D-galactose-treated mice (Lu et al., 2007).

Depression

The α/β -amyrin, a triterpene isomeric mixture from *Protium heptaphyllum*, has been shown to decrease the immobility time in the behavior despair test in mice (Yildiz et al., 2002). In another study, β -amyrin palmitate showed tonic inhibitory action on depression. Further, it was suggested that the release of norepinephrine from newly synthesized pools might account for the antidepressant actions of β -amyrin palmitate (Chen et al., 2003; Subarnas et al., 1993).

Convulsions

The distinct GABA_A-receptor related properties of lupane type triterpenoids such as betulin, betulinic acid and lupeol have been reported *in vivo* and *in vitro*. Muceniece et al. (2008) has showed that betulin competed with [3H]GABA for binding to the corresponding sites on the GABA_A receptor, whereas betulinic acid and lupeol did not show any binding affinity. Further, antagonistic action of betulin against bicuculline, a convulsant drug, following central and peripheral administration of betulin has been reveled by this study.

Pain and nociception

Various triterpenoids such as lupeol, tormentic acid, betulin, betulinic acid and *epi*betulin have been reported to produce significant antinociceptive and/or anti-inflammatory

Phytopharmacology 2013, 4(2), 354-372

activities by different mechanisms such as inhibition of cyclo and lipoxygenase pathways (Calixto et al., 2000, Moroney et al., 1988). Tormentic acid, a naturally occurring pentacyclic triterpenoid found in a variety of plants, has been shown to possess anti-allodynic action in two different models of chronic pain in mice: neuropathic pain caused by partial constriction of the sciatic nerve and inflammatory pain caused by plantar injection of complete Freund's adjuvant (Bortalanza et al., 2002).

Anti-nociceptive effect of lupeol isolated from *Zanthoxylum rhoifolium* in models of acute pain in rodents has been evaluated. In this study, lupeol reduced the glutamate-evoked nociceptive response and ameliorated the neurogenic nociception induced by intraplantar injection of capsaicin that stimulated nerve endings causing intense thermal and nociceptive pain (Sakurada et al., 1992, Santos and Calixto, 1997).

The lupane triterpenoids such as betulin, 28-acetoxy-betulin, *epi*betulin, *epi*betulinic acid, and betulonic acid have been shown to possess potent anti-inflammatory activity through inhibition of nitric oxide and prostaglandin E_2 production in mouse macrophages stimulated with bacterial endotoxin (Misko et al,1993, Moroney et al.,1988). Luiz et al. (2007) has evaluated the ethanolic extract from roots of *Humirianthera ampla* against neurogenic and inflammatory models of nociception in experimental animals and concluded that the nociceptive activity might be attributed to di- and triterpenoids, the chief constituents of *H. ampla*.

Oleanolic acid, the anti-inflammatory pentacyclic triterpenoid, produced tonic inhibitory effects on capsaicin-evoked acute nociception due to mechanisms possibly involving endogenous opioids, nitric oxide, and potassium-ATP-channel opening (Maia et al., 2006) in experimental animals. The anti-inflammatory mechanisms of the ursane type pentacyclic triterpenoid, β -boswellic acid and its derivatives have been studied by Giner et al. (2000). The study concluded that β -boswellic acid and its derivatives produced anti-inflammatory action through inhibition of 5-lipoxygenase.

Anxiety

The oral and intraperitoneal administration of betulinic acid produced anti-anxiety activity in animals. Further, a pharmaceutical composition containing betulinic acid has been patented as a means for prevention or treatment of anxiety (Durst et al., 2002).

The anxiolytic effects of the mixture of α/β -amyrin, the pentacyclic triterpenoids isolated from the stem bark resin of *Protium heptaphyllum*, has been demonstrated by different animal models. The results of this study showed that α/β -amyrin significantly decreased the number of crossings, grooming, rearing and the time of permanence and the number of entrances in the close arms whereas, increased the time of permanence and the number of entrances in the open arms (Dias et al., 2005; Herrera-Ruiz et al., 2006; Morris et al., 2006).

Sedation and hypnotics

 α/β -amyrin has also been evaluated for their sedative effects in mice (Petty, 1995). Subarnas et al. (1993) have investigated hypnotic potential of β -amyrin palmitate in pentobarbitone-induced narcosis in mice and concluded that administration of β -amyrin palmitate potentiated the pentobarbitone-provoked narcosis in experimental animals.

Biological role of tetracyclic triterpenes

Pain and nociception

A number of studies indicate that tetracyclic triterpenes play a central role in the manageeement of pain. Scott et al. (2004) have investigated the anti-nociceptive activity of euphol against nociceptive response induced by ligation of the sciatic nerve and injection of complete Freund's adjuvant in rat paw and spinal cord. The results showed significant effect of euphol in preventing the mechanical nociception induced by sciatic nerve ligation and also ameliorated the levels and mRNA of cytokines in both paw and spinal cord tissues following injection of complete Freund's adjuvant. An interesting finding of this study was that even in higher doses, euphol did not cause any relevant action in the central nervous system. Recently, Dutra et al. (2011 & 2012) evaluated the molecular mechanism of anti-nociceptive properties of euphol and found that anti-nociception effect of euphol was related with its ability to inhibit the activation and/or release of various inflammatory mediators such as IL-1 β , IL-6 and TNF- α , as well as due to the blockade of neutrophils influx in the rat paw and spinal cord tissue, respectively.

The analgesic effects of the extract of roots and tubers of Wilbrandia ebracteata has been studied by Peters et al. (1997). In this study, it was suggested that cucurbitacins, the chief constituents of the W. ebracteata extracts, might be responsible for the analgesic activity by inhibition of PGE2 production. Similarly, Bralley et al. (2007) have evaluated topical application of tirucallol, a tetracyclic triterpene from Euphorbia lacteal latex, against mouse model of ear oedema. The results from this study showed that tirucallol improved ear oedema and inhibited the influx of polymorphonuclear cells to inflammed area of mouse ear. In another study, the effect of tirucallol on some macrophage functions has been analyzed in vitro (Carlson et al. 1989). It was postulated that non-toxic concentrations of tirucallol inhibited nitrite production in lipopolysaccharide-stimulated macrophages. Anti-nociceptive and antiinflammatory effects of (-)-cassine, a tetracyclic triterpene from Senna spectabilis, has been evaluated using pharmacological, behavioural and biochemical approaches in experimental animals. The study indicated that pretreatment with (-)-cassine reduced carrageenan-induced mechanical and thermal nociception, and prostaglandin E2-, Freund's complete adjuvant-, (IL)-1b-, (IL)-6- and keratinocyte-derived chemokine-provoked hyperalgesia (Kathryn et al., 2012). De Souza et al. (2009) have reported potent antinociceptive property of filicene, a triterpene from Adiantum cuneatum leaves, in acetic acid-induced writhing, capsaicin and glutamate-induced nociception models in mice. Li et al. (2012) has investigated the antinociceptive effects of escin against formalin-induced activation of c-Fos and phosphorylated p38 MAPK in the rat spinal cord. In this study, escin decreased pain-related behaviours, c-Fos and phosphorylated p38 MAPK expressions. In another study, the molecular mechanisms underlying antinociceptive effect of the 3β, 6β, 16β-trihydroxylup-20(29)-ene (TTHL), a triterpene, in mice was evaluated. The findings of this study suggested that TTHL produced antinociceptive effect that was dependent on opioid and serotonergic systems, Gi/o protein activation and the opening of specific K+ channels (Longhi-Balbinot et al., 2009).

Anxiety

An infusion prepared with aerial parts from *Galphimia glauca* has been widely used in Mexican traditional medicine as a remedy for nervous excitement. The sedative activity of a methanolic extract from this plant has been demonstrated by neuropharmacological tests such as elevated plus-maze, light-dark test and the forced swimming models. The effect was attributed to the nor-secotriterpene named galphimine B (Herrera-Ruiz et al., 2006). Similarly, galphimine-B has been investigated for anxiolytic effect by means of a double blind clinical trial. It was demonstrated that galphimine-B could be a promising therapeutic candidates for the patients with generalized anxiety (Jiménez-Ferrer et al., 2011).

Others

Neuroprotective activities

Asiatic acid has been studied for its neuroprotective effects *in vitro* and *in vivo*. Vogel et al. (1990) showed that asiatic acid produced neuroprotection through reduction in apoptotic cell death, neutralization of reactive oxygen species and stabilizing the mitochondrial membrane potential. The study also showed that oral administration of asiatic acid markedly improved Morris water maze test experience, reduced lipid peroxidation levels and normalized glutathione and SOD levels in the rat brain. *Ganoderma lucidum*, the rich source of triterpenoids, is one of the most popular medicinal fungi with a long history of use in Asian countries (Sanodiya et al., 2009). Chen et al. (2012) has investigated cerebroprotective activity of *G. lucidum* against mitochondrial toxins in mouse model of Huntington's disease and suggested that the effect could be attributed to the triterpenoids from this plant. Similarly, ursolic acid, oleanolic acid, 3-*epi*ursolic, 3-*epi*oleanolic acid and derivatives of ursolic acid and oleanolic acid are abundantly present in *Verbena officinalis* (Deepak and Handa, 1998). The study of Lai et al. (2006) indicated that triterpenoids could play a central role in the effect of *V. officinalis* on β -amyloid peptide-induced cytotoxicity.

Diverse efforts have been made to discover anti-Alzheimer's agents from natural sources. Various group of chemicals like ginsenosides, gingkolides, and canabinoids as potential anti- Alzheimer's disease agents have been studied. These compounds exhibit promising in vitro and in vivo biological activities, but are still to be tested clinically. Other compounds such as cornel iridoid glycoside, oleanolic acid, tenuifolin, cryptotanshinone and ursolic acid have outstanding neuroprotective effects in in vitro assays. These compounds can exert beneficial effects on central nervous system directly or indirectly by acting on peripheral targets (Yoo and Park, 2012). Ginsenosides enjoy a special attention among triterpenoids when it comes to pharmacological activities. Ginsenosides have been research targets over the last three decades to explain ginseng actions and a wealth of literature has been presented reporting on ginsenosides' effects on the human body. Recently, there is increasing evidence on beneficial effects of ginsenosides to the central nervous system. Using a wide range of in vitro and in vivo models, researchers have attributed these effects to specific pharmacological actions of ginsenosides on cerebral metabolism, oxidative stress and radical formation, neurotransmitter imbalance and membrane stabilizing effects, and even antiapoptotic effects. Modulating these particular mechanisms by ginsenosides has thus been reported to exert either general stimulatory effects on the brain functions or protecting the CNS against various disease conditions (Radad et al., 2011). Another study reported that maslinic acid exerts antiapoptotic as well as neuroprotective effect through inhibition of inducible nitic oxide synthase and normalization of caspase expression/activation against oxygen-glucose deprivationinduced neuron damage (Qian et al., 2011). Considerable evidence has been accumulated demonstrating an important role for inflammation in ischemic brain injury and its contribution to greater cerebral damage after ischemia. Blocking the inflammatory reaction promotes neuroprotection and shows therapeutic potential for clinical treatment of ischemic brain injury (Brea et al., 2009; Dos-Anjos et al., 2009). Moreover, celastrol a pentacyclic triterpenoid has been showed to possess anti-ischemic action against ischemia-reperfusion-induced cerebral injury through downregulation of the expression of p-JNK, p-c-Jun and NF-kB (Lia et al., 2012). This ameliorating effect of celastrol on the expression of p-JNK, p-c-Jun and NF- κ B is evidenced by the another set of experiments where celastrol has been showed to possess anti-inflammatory and anti-apoptotic activities by inhibition of NF-kB activation and MAPK inactivation including JNK (Kannaiyan et al., 2011a, 2011b; Kim et al., 2009). Ursolic acid, a natural pentacyclic triterpenoid acid, a well-known anti-oxidative and antiinflammatory reagent, protects the brain against ischemic injury by promoting the activation of nuclear factor-erythroid 2-related factor 2 pathway and downregulation of the expression of TLR4 and NF-kB (Li et al., 2012). Escin, a natural mixture of triterpenoid saponin has been showed to improve learning and memory recovery and reduce hippocampal damage in the cerebral ischemic mice (Zhang et al., 2010). This study reported that anti-ischemic action conferred by escin was by downregulation of certain inflammatory gene expression and upregulating the expression of granulocyte-macrophage colony-stimulating factor in experimental animals.

In conclusion, triterpenes comprise one of the most interesting groups of natural products due to their high potential as pharmacological agents. Triterpenes are ubiquitously present in plants of ethnomedicinal use. Many such compounds can either be used directly as active compounds or modified to increase their selectivity and potency. Although they have generally been examined for their anti-inflammatory and antiviral properties, their possible use as immunosuppressant drugs should be considered for future research. In addition, new paths of investigation should be pursued, including studies on their effects on transcriptional pathways as well as their implication in immune responses. Several structural groups of triterpenes have demonstrated specificity against transcriptional factors which could be of particular interest in treating inflammation, cancer, and immune diseases.

Conflict of interest

None of the authors have any conflict of interest to declare.

References

- Abe I, Rohmer M, Prestwich GD. (1993). Enzymatic cyclization of squalene and oxidosqualene to sterols and triterpenes. *Chemical Reviews* 93, 2189-2206.
- Afaq F, Saleem M, Aziz MH, Mukhtar H. (2004). Inhibition of 12-O-tetradecanoylphorbol-13acetate-induced tumor promotion markers in CD-1 mouse skin by oleandrin. *Toxicology and Applied Pharmacology* 195, 361-369.
- Aggarwal BB, Shishodia S. (2006). Molecular targets of dietary agents for prevention and therapy of cancer. *Biochemistry and Pharmacology* 71, 1397-1421.

- Bortalanza LB, Ferreira J, Hess SC, Monache FD, Yunes RA, Calixto JB. (2002). Anti-allodynic action of the tormentic acid, a triterpene isolated from plant, against neuropathic and inflammatory persistent pain in mice. *European Journal of Pharmacology* 453, 203-208.
- Bralley EE, Hargrove JL, Greenspan P, Hartle DK. (2007). Topical anti-inflammatory activities of *Vitis rotundifolia* (Muscadine grape) extracts in the tetradecanoylphorbol acetate model of ear inflammation. *Journal of Medicinal Food* 4, 636-642.
- Brea D, Sobrino T, Ramos-Cabrer P, Castillo J. (2009). Inflammatory and neuroimmunomodulatory changes in acute cerebral ischemia. *Cerebrovascular Disease* 27, 48-64.
- Buckingham J. (2002). Dictionary of Natural Products, Web version. Chapman & Hall/CRC Press.
- Calixto JB, Beirith A, Ferreira J, Santos AR, Cechinel FV, Yunes RA. (2000). Naturally occurring antinociceptive substances from plants. *Phytotherapy Research* 14, 401-418.
- Carlson RP, O'Neil-Davis L, Calhoun W, Datko L, Musser JH, Kreft AF, Chang JY. (1989). Effect of a 5-LO/cycloxygenase (CO) inhibitor, WY-47, 288 on cutaneous models of inflammation. *Agents Actions* 26, 319-328.
- Chan KT, Li K, Liu SL, Chu KH, Toh M, Xie WD. (2010). Cucurbitacin B inhibits STAT3 and the Raf/MEK/ERK pathway in leukemia cell line K562. *Cancer Letters* 289, 46-52.
- Chappell J. (1995). Biochemistry and molecular biology of the isoprenoid biosynthetic pathway in plants. *Annual Review of Plant Physiology and Plant Molecular Biology* 46, 521-547.
- Chen LW, Horng LY, Wu CL, Sung HC, Wu RT. (2012). Activating mitochondrial regulator PGC-1α expression by astrocytic NGF is a therapeutic strategy for Huntington's disease. *Neuropharmacology* 63, 719-732.
- Chen NH, Liu JW, Zhong JJ. (2008). Ganoderic acid Me inhibits tumor invasion through downregulating matrix metalloproteinases 2/9 gene expression. *Journal of Pharmacological Sciences* 108, 212-216.
- Chen Y, Han T, Qin L, Rui Y, Zheng H. (2003). Effect of total triterpenes from *Centella asiatica* on the depression behavior and concentration of amino acid in forced swimming test. *Zhong Yao Cai* 26, 870-873.
- Chen Z, Wu Q, Chen Y, He J. (2008). Effects of betulinic acid on proliferation and apoptosis in Jurkat cells and its *in vitro* mechanism. *Journal of Huazhong University of Science and Technology Medical Sciences* 28, 634-638.
- Chintharlapalli S, Papineni S, Ramaiah SK, Safe S. (2007). Betulinic acid inhibits prostate cancer growth through inhibition of specificity protein transcription factors. *Cancer Research* 67, 2816-2823.
- Chu R, Zhao C, Griffin C, Staub RE, Shoemaker M, Climent J, Leitman D, Cohen I, Shtivelman E, Fong S. (2010). Selective concomitant inhibition of mTORC1 and mTORC2 activity in estrogen receptor negative breast cancer cells by BN107 and oleanolic acid. *International Journal of Cancer* 127, 1209-1219.
- Connolly JD, Hill RA. (2001). Triterpenoids. Natural Product Reports 18, 560-578.
- De Souza MM, Pereira MA, Ardenghi JV, Mora TC, Bresciani LF, Yunes RA, Monache FD, Cechinel-Filho V. (2009). Filicene obtained from *Adiantum cuneatum* interacts with the cholinergic, dopaminergic, glutamatergic, GABAergic, and tachykinergic systems to exert antinociceptive effect in mice. *Pharmacology, Biochemistry and Behavior* 93, 40-46.
- Deeb D, Gao X, Dulchavsky SA, Gautam SC. (2008). CDDO-Me inhibits proliferation, induces apoptosis, down-regulates Akt, mTOR, NF-kappaB and NF-kappaB-regulated antiapoptotic and proangiogenic proteins in TRAMP prostate cancer cells. *Journal of Experimental Therapeutics and Oncology* 7, 31-39.
- Deepak M, Handa SS. (1998). 3a,24-dihydroxy-urs-12-en-28-oic acid from Verbena officinalis. Phytochemistry 49, 269-271.
- Dias R, Sheppard WF, Fradley RL, Garret EM, Stanley LJ, Tye SJ. (2005). Evidence for a significant role of alpha 3-containing GABA-A receptors in mediating the anxiolytic effects of benzod-iazepines. *Journal of Neuroscience* 25, 10682-10688.

- Dos-Anjos S, Martı'nez-Villayandre B, Montori S, Regueiro-Purrin^oos MM, Gonzalo-Orden JM, Ferna'ndez-Lo'pez A. (2009). Global ischemia-induced modifications in the expression of AMPA receptors and inflammation in rat brain. *Brain Research* 1287, 20-27.
- Durst T, Merali Z, Arnason JT, Sanchez-Vindas EP, Poveda AL. (2002). Anxiolytic Marcgraviaceae compositions containing betulinic acid, betulinic acid derivates, and methods. WO/2002/091858.
- Dutra RC, Claudino RF, Bento AF, Marcon R, Schmidt EC, Bouzon ZL, Pianowski LF, Calixto JB. (2011). Preventive and therapeutic euphol treatment attenuates experimental colitis in mice. *PLoS ONE* 6, e27122.
- Dutra RC, Claudino RF, Bento AF, Marcon R, Schmidt EC, Bouzon ZL, Pianowski LF, Calixto JB. (2012). Euphol prevents experimental autoimmune encephalomyelitis in mice: evidence for the underlying mechanisms. *Biochemical Pharmacology* 83, 531-542.
- Eggersdofer M. (2005). Terpenes In: Ullmann's encyclopedia of industrial chemistry, electronic release. Weinheim:Wiley-VCH.
- Fernandez MA, de las Heras B, Garcia MD, Saenz MT, Villar A. (2001). New insights into the mechanism of action of the anti-inflammatory triterpene lupeol. *The Journal of Pharmacy and Pharmacology* 53, 1533-1539.
- Fonranay S, Grare M, Mayer J, Finance C, Duval RE. (2008). Ursolic, oleanolic and betulinic acids: antibacterial spectra and selectivity indexes. *Journal of Ethnopharmacology* 120, 272-276.
- Fulda S, Friesen C, Los M, Scaffidi C, Mier W, Benedict M, Nunez G, Krammer PH, Peter ME, Debatin KM. (1997). Betulinic acid triggers CD95 (APO-1/Fas)-and p53-independent apoptosis via activation of caspases in neuroectodermal tumors. *Cancer Research* 57, 4956-4964.
- Gershenzon J, Kreis W. (1999). Biosynthesis of monoterpenes, sesquiterpenes, diterpenes, sterols, cardiac glycosides and steroid saponins, In: Biochemistry of plant secondary metabolites. Annual Plant Reviews; Wink, M. (Ed.), Sheffield Academic Press: Sheffield, UK, 2, pp. 222-299.
- Gildemeister E, Hoffmann F. (1960). Die ätherischen Öle, 1-7, 4th ed. Berlin: Akademie Verlag.
- Giner RM, Villalba ML, Recio MC, Máñez S, Cerdá-Nicolás M, Ríos JL. (2000). Anti-inflammatory glycoterpenoids from *Scrophularia auriculata*. *European Journal of Pharmacology* 389, 243-252.
- Gunatilaka AAL. (1986). Triterpenoids and steroids of Sri Lankan plants: a review of occurance and chemistry. *Journal of National Science and Country* 14, 1-54.
- Gurfinkel DM, Chow S, Hurren R, Gronda M, Henderson C, Berube C, Hedley DW, Schimmer AD. (2006). Disruption of the endoplasmic reticulum and increases in cytoplasmic calcium are early events in cell death induced by the natural triterpenoid asiatic acid. *Apoptosis* 11, 1463-1471.
- Hanausek M, Ganesh P, Walaszek Z, Arntzen CJ, Slaga TJ, Gutterman JU. (2001). Avicins, a family of triterpenoid saponins from *Acacia victoriae* (Bentham), suppress H-ras mutations and aneuploidy in a murine skin carcinogenesis model. *Proceedings of the National Academy of Science of United State of America* 98, 11551-11556.
- Hanson JR. (2003). Natural products: The secondary metabolites. The Royal Society of Chemistry: Cambridge, UK, pp. 112-121.
- Haralampidis K, Trojanowska M, Osbourn AE. (2002). Biosynthesis of triterpenoid saponins in plants. In: Scheper, T. (Ed.), Springer Verlag: Berlin, Heidelberg, Germany. Advances in Biochemical Engineering/Biotechnology 75, 32-49.
- Haridas V, Nishimura G, Xu ZX, Connolly F, Hanausek M, Walaszek Z, Zoltaszek R, Gutterman JU. (2009). Avicin D: a protein reactive plant isoprenoid dephosphorylates Stat 3 by regulating both kinase and phosphatase activities. *PLoS One* 4, e5578.

- Harikumar KB, Sung B, Pandey MK, Guha S, Krishnan S, Aggarwal BB. (2010). Escin, a pentacyclic triterpene, chemosensitizes human tumor cells through inhibition of NF-{kappa}B signaling pathway. *Molecular Pharmacology* 77, 818-827.
- He MF, Liu L, Ge W, Shaw PC, Jiang R, Wu LW, But PP. (2009). Antiangiogenic activity of *Tripterygium wilfordii* and its terpenoids. *Journal of Ethnopharmacology* 121, 61-68.
- Herrera-Ruiza M, Jime'nez-Ferrera JE, De Limab TCM, Avile's-Montesc D, Pe'rez-Garci D, Gonza' lez-Cortazara D, Tortorielloa J. (2006). Anxiolytic and antidepressant-like activity of a standardized extract from *Galphimia glauca*. *Phytomedicine* 13, 23-28.
- Honda T, Liby KT, Su X, Sundararajan C, Honda Y, Suh N, Risingsong R, Williams CR, Royce DB, Sporn MB, Gribble GW. (2006). Design, synthesis, and anti-inflammatory activity both *in vitro* and *in vivo* of new betulinic acid analogues having an enone functionality in ring A. *Bioorganic and Medicinal Chemistry Letters* 16, 6306-6309.
- Hsu YL, Kuo PL, Chiang LC, Lin CC. (2004). Involvement of p53, nuclear factor kappaB and Fas/Fas ligand in induction of apoptosis and cell cycle arrest by saikosaponin d in human hepatoma cell lines. *Cancer Letters* 213, 213-221.
- Hu XM, Zhang Y, Zeng FD. (2004). Effects of beta-aescin on apoptosis induced by transient focal cerebral ischemia in rats. *Acta Pharmacologica Sinica* 25, 1267-1275.
- Hwang YH, Lee JW, Hahm ER, Jung KC, Lee JH, Park CH, Rhee HS, Ryu JM, Kim HK, Yang CH. (2005). Momordin I, an inhibitor of AP-1, suppressed osteoclastogenesis through inhibition of NF-kappaB and AP-1 and also reduced osteoclast activity and survival. *Biochemical and Biophysical Research Communication* 337, 815-823.
- Jang SM, Yee ST, Choi J, Choi MS, Do GM, Jeon SM, Yeo J, Kim MJ, Seo KI, Lee MK. (2009). Ursolic acid enhances the cellular immune system and pancreatic β-cell function in streptozotocin-induced diabetic mice fed a high fat diet. *International Immunopharmacology* 9, 113-119.
- Jenner H, Townsend B, Osbourn A. (2005). Unraveling triterpene glycoside synthesis in plants: phytochemistry and functional genomics join forces. *Planta* 220, 503-506.
- Jeremias I, Steiner HH, Benner A, Debatin KM, Herold-Mende C. (2004). Cell death induction by betulinic acid, ceramide and TRAIL in primary glioblastoma multiforme cells. *Acta Neurochirurgica* (Wien) 146, 721-729.
- Jiang J, Grieb B, Thyagarajan A, Sliva D. (2008). Ganoderic acids suppress growth and invasive behavior of breast cancer cells by modulating AP-1 and NF-kappaB signaling. *International Journal of Molecular Medicine* 21, 577-584.
- Jiménez-Ferrera E, Herrera-Ruiza M, Ramírez-Garcíaa R, Herrera-Arellanoa A, Tortoriello J. (2011). Interaction of the natural anxiolytic galphimine-B with serotonergic drugs on dorsal hippocampus in rats. *Journal of Ethnopharmacology* 137, 724-729.
- Jung HW, Chung YS, Kim YS, Park YK. (2007). Celastrol inhibits production of nitric oxide and pro-inflammatory cytokines through MAPK signal transduction and NF-kappa B in LPSstimulated BV-2 microglial cells. *Experimental and Molecular Medicine* 39, 715-721.
- Kannaiyan R, Shanmugam MK, Sethi G. (2011a). Molecular targets of celastrol derived from thunder of god vine: potential role in the treatment of inflammatory disorders and cancer. *Cancer Letters* 303, 9-20.
- Kannaiyan R, Manu KA, Chen L, et al. (2011b). Celastrol inhibits tumor cell proliferation and promotes apoptosis through the activation of c-Jun N-terminal kinase and suppression of PI3 K/Akt signaling pathways. *Apoptosis* 16, 1028-1041.
- Kathryn ABS, Silva D, Manjavachi MN, Paszcuk AF, Pivatto M, Viegas C, Bolzani VS, Calixto JB. (2012). Plant derived alkaloid cassine induces anti-inflammatory and anti-hyperalgesics effects in both acute and chronic inflammatory and neuropathic pain models. *Neuropharmacology* 62, 967-977.

- Kim DY, Park JW, Jeoung D, Ro JY. (2009). Celastrol suppresses allergen-induced airway inflammation in a mouse allergic asthma model. *European Journal of Pharmacology* 612, 98-105.
- Kim JH, Ju EM, Lee DK, Hwang HJ. (2002). Induction of apoptosis by momordin I in promyelocytic leukemia (HL-60) cells. *Anticancer Research* 22, 1885-1889.
- Kunnumakkara AB, Nair AS, Sung B, Pandey MK, Aggarwal BB. (2009). Boswellic acid blocks signal transducers and activators of transcription 3 signaling, proliferation, and survival of multiple myeloma via the protein tyrosine phosphatase SHP-1. *Molecular Cancer Research* 7, 118-128.
- Lai SW, Yu MS, Yuen WH, Chang RC. (2006). Novel neuroprotective effects of the aqueous extracts from *Verbena officinalis* Linn. *Neuropharmacology* 50, 641-650.
- Lee JS, Miyashiro H, Nakamura N, Hattori M. (2008a). Two new triterpenes from the rhizome of *Dryopteris crassirhizoma*, and inhibitory activities of its constituents on human immunodeficiency virus-1 protease. *Chemical and Pharmaceutical Bulletin* 56, 711-714.
- Lee SU, Park SJ, Kwak HB, Oh J, Min YK, Kim SH. (2008b). Anabolic activity of ursolic acid in bone: stimulating osteoblast differentiation *in vitro* and inducing new bone formation *in vivo*. *Pharmacological Research* 58, 290-296.
- Lee TK, Poon RT, Wo JY, Ma S, Guan XY, Myers JN, Altevogt P, Yuen AP. (2007). Lupeol suppresses cisplatin-induced nuclear factor-kappa B activation in head and neck squamous cell carcinoma and inhibits local invasion and nodal metastasis in an orthotopic nude mouse model. *Cancer Research* 67, 8800-8809.
- Li L, Zhang X, Cui L, Wang L, Liu H, Ji H, Du Y. (2012). Ursolic acid promotes the neuroprotection by activating Nrf2 pathway after cerebral ischemia in mice. *Brain Research* 25, 2497-2499.
- Li Q, Ouyang H, Wang P, Zeng W. (2012). The anti-nociceptive effect of intrathecal escin in the rat formalin test. *European Journal of Pharmacology* 674, 234-238.
- Longhi-Balbinot DT, Pietrovski EF, Gadotti VM, Martins DF, Facundo VA, Santos ARS. (2009). Spinal antinociception evoked by the triterpene 3β, 6β, 16β-trihydroxylup-20(29)-ene in mice: evidence for the involvement of the glutamatergic system via NMDA and metabotropic glutamate receptors. *European Journal of Pharmacology* 623, 30-36.
- Lu J, Zheng Y, Wu D, Luo L, Sun D, Shan Q. (2007). Ursolic acid ameliorates cognition deficits and attenuates oxidative damage in the brain of senescent mice induced by D-galactose. *Biochemical Pharmacology* 74, 1078-1090.
- Lui VW, Yau DM, Wong EY, Ng YK, Lau CP, Ho Y, Chan JP, Hong B, Ho K, Cheung CS, Tsang CM, Tsao SW, Chan AT. (2009). Cucurbitacin I elicits anoikis sensitization, inhibits cellular invasion and *in vivo* tumor formation ability of nasopharyngeal carcinoma cells. *Carcinogenesis* 30, 2085-2094.
- Luiz AP, Moura J, Meotti FC, Guginski G, Guimar aes CLS, Azevedo MS, Rodrigues ALS, Santos ARS. (2007). Antinociceptive action of ethanolic extract obtained from roots of *Humirian-thera ampla* Miers. *Journal of Ethnopharmacology* 114, 355-363.
- Ma CM, Cai SQ, Cui JR, Wang RQ, Tu PF, Hattori M, Daneshtalab M. (2005). The cytotoxic activity of ursolic acid derivatives. *European Journal of Medicinal Chemistry* 40, 582-589.
- Maia JL, Lima-J'unior RCP, Melo CM, David JP, David JM, Campos AR, Santos FA, Rao VSN. (2006). Oleanolic acid, a pentacyclic triterpene attenuates capsaicin-induced nociception in mice: possible mechanisms. *Pharmacological Research* 54, 282-286.
- Mangas S, Bonfill M, Osuna L, Moyano E, Tortoriello J, Cusido RM, Pinol MT, Palaźom J. (2006). The effect of methyl jasmonate on triterpene and steriol metabolisms of *Centella asiatica*, *Ruscus aculeatus* and *Galphimia glauca* cultured plants. *Phytochemistry* 67, 2041-2049.
- Manna SK, Sah NK, Newman RA, Cisneros A, Aggarwal BB. (2000). Oleandrin suppresses activateion of nuclear transcription factor-kappa B, activator protein-1, and c-Jun NH2-terminal kinase. *Cancer Research* 60, 3838-3847.

- Manu KA, Kuttan G. (2008). Ursolic acid induces apoptosis by activating p53 and caspase-3 gene expressions and suppressing NF-kappaB mediated activation of bcl-2 in B16F-10 melanoma cells. *International Immunopharmacology* 8, 974-981.
- Matsuda H, Li Y, Murakami T, Ninomiya K, Yamahara J, Yoshikawa M. (1997). Effects of escins Ia, Ib, IIa, and IIb from horse chestnut, the seeds of *Aesculus hippocastanum* L. on acute inflammation in animals. *Biological and Pharmaceutical Bulletin* 20, 1092-1095.
- Matsuda H, Murakami T, Li Y, Yamahara J, Yoshikawa M. (1998). Mode of action of escins Ia and IIa and E, Z-senegin II on glucose absorption in gastrointestinal tract. *Bioorganic and Medicinal Chemistry* 6, 1019-1023.

McGarvey DJ, Croteau R. (1997). Terpenoid metabolism. Plant Cell 7, e26.

- Medeiros R, Otuki MF, Avellar MC, Calixto JB. (2007). Mechanisms underlying the inhibitory actions of the pentacyclic triterpene alpha-amyrin in the mouse skin inflammation induced by phorbol ester 12-O-tetradecanoylphorbol-13-acetate. *European Journal of Pharmacology* 559, 227-235.
- Min SW, Jung SH, Cho KH, Kim DH. (2008). Antihyperlipidemic effects of red ginseng, *Crataegii Fructus* and their main constituents ginsenoside Rg3 and ursolic acid in mice. *Biomolecular Therapy* 16, 364-369.
- Misko TP, Schilling RJ, Salvemini D, Moore WM, Currie MG. (1993). Activity of lupane triterpenoids from Maytenus species. *Analytical Biochemistry* 214, 11.
- Montopoli M, Froldi MC, Comelli M, Prosdocimi L. (2007). Caparrotta, aescin protection of human vascular endothelial cells exposed to cobalt chloride mimicked hypoxia and inflammatory stimuli. *Planta Medica* 73, 285-288.
- Moroney MA, Alcaraz MJ, Forder RA, Carey F, Hoult JRS. (1988). Activity of lupane triterpenoids from Maytenus species. *Journal of Pharmacy and Pharmacology* 40, 787.
- Morris HV, Dawson GR, Reynolds DS, Atack JR, Stephens DN. (2006). Both alpha 2 and alpha 3 GABA-A receptor subtypes mediate the anxiolytic properties of benzodiazepine site ligands in the conditioned emotional response paradigm. *European Journal of Neuroscience* 23, 2495-2504.
- Muceniece R, Saleniece K, Rumaks J, Krigere L, Dzirkale Z, Mezhapuke R, Zharkova O, Klusa V. (2008). Betulin binds to γ-aminobutyric acid receptors and exerts anticonvulsant action in mice. *Pharmacology, Biochemistry and Behavior* 90, 712-716.
- Murtaza I, Saleem M, Adhami VM, Hafeez BB, Mukhtar H. (2009). Suppression of cFLIP by lupeol, a dietary triterpene, is sufficient to overcome resistance to TRAIL-mediated apoptosis in chemoresistant human pancreatic cancer cells. *Cancer Research* 69, 1156-1165.
- Nasira MN, Habsahd M, Zamzuria I, Rammese G, Hasnanc J, Abdullah J. (2011). Effects of asiatic acid on passive and active avoidance task in male Spraque-Dawley rats. *Journal of Ethnopharmacology* 134, 203-209.
- Pang X, Yi Z, Zhang J, Lu B, Sung B, Qu W, Agrawal BB, Liu M. (2010). Celastrol suppresses angiogenesis-mediated tumor growth through inhibition of AKT/mammalian target of rapamycin pathway. *Cancer Research* 70, 1951-1959.
- Park BC, Bosire KO, Lee ES, Lee YS, Kim JA. (2005). Asiatic acid induces apoptosis in SK-MEL-2 human melanoma cells. *Cancer Letters* 218, 81-90.
- Park BC, Paek SH, Lee YS, Kim SJ, Lee ES, Choi HG, Yong CS, Kim JA. (2007). Inhibitory effects of asiatic acid on 7,12-dimethylbenz[a]anthracene and 12-*O*-tetradecanoylphorbol 13-acetate-induced tumor promotion in mice. *Biological and Pharmaceutical Bulletin* 30, 176-179.
- Peters RR, Farias MR, Ribeiro do Vale PM. (1997). Anti-inflammatory and analgesic effects of cucurbitacins from *Wilbrandia ebracteata*. *Planta Medica* 63, 525-528.
- Petty F. (1995). GABA and mood disorders: A brief review and hypothesis. *Journal of Affective Disorders* 34, 275-281.
- Phillips DR, Rasberry JM, Bartel B, Matsuda SPT. (2006). Biosynthestic diversity in plant triterpene cyclization. *Current Opinion in Plant Biology* 9, 305-314.

- Prasad S, Madan E, Nigam N, Roy P, George J, Shukla Y. (2009). Induction of apoptosis by lupeol in human epidermoid carcinoma A431 cells through regulation of mitochondrial, Akt/PKB and NF-kappa-B signaling pathways. *Cancer Biology and Therapy* 8, 1632-1639.
- Qian Y, Guan T, Tang X, Huang L, Huang M, Li Y, Sun H. (2011). Maslinic acid, a natural triterpenoid compound from *Olea europaea*, protects cortical neurons against oxygen-glucose deprivation-induced injury. *European Journal of Pharmacology* 670, 148-153.
- Radad K, Moldzio R, Rausch WD. (2011). Ginsenosides and their CNS targets. CNS Neuroscience and Therapeutics 17, 761-768.
- Ramachandran S, Prasad NR. (2008). Effect of ursolic acid, a triterpenoid antioxidant, on ultraviolet-B radiation-induced cytotoxicity, lipid peroxidation and DNA damage in human lymphocytes. *Chemico-Biological Interactions* 176, 99-107.
- Raphael TJ, Kuttan G. (2003). Effect of naturally occurring triterpenoids glycyrrhizic acid, ursolic acid, oleanolic acid and nomilin on the immune system. *Phytomedicine* 10, 483-489.
- Resende FA, de Andrade Barcala CAM, da Silva Faria MC, Kato FH, Cunha WR, Tavares DC. (2006). Antimutagenicity of ursolic acid and oleanolic acid against doxorubicin-induced clastogenesis in balb/c mice. *Life Sciences* 79, 1268-1273.
- Rothkopf M, Vogel G. (1976). New findings on the efficacy and mode of action of the horse chestnut saponin escin. *Arzneimittelforschung* 26, 225-235.
- Sailer ER, Subramanian LR, Rall B, Hoernlein RF, Ammon HP, Safayhi H. (1996). Acetyl-11-ketobeta-boswellic acid (AKBA): structure requirements for binding and 5-lipoxygenase inhibitory activity. *British Journal of Pharmacology* 117, 615-618.
- Sakurada T, Katsumata K, Tanno K, Sakurada S, Kisara K. (1992). The capsaicin test in mice for evaluating tachykinin antagonists in the spinal cord. *Neuropharmacology* 31, 1279-1285.
- Saleem M. (2009). Lupeol, a novel anti-inflammatory and anti-cancer dietary triterpene. *Cancer Letters* 285, 109-115.
- Salminen A, Lehtonen M, Paimela T, Kaarniranta K. (2010). Celastrol: molecular targets of Thunder God Vine. *Biochemical and Biophysical Research Communications* 394, 439-442.
- Sandermann W. (1960). Naturharze, Terpentinöl, Tallöl. Berlin: Springer Verlag.
- Sanodiya BS, Thakur GS, Baghel RK, Prasad GB, Bisen PS. (2009). *Ganoderma lucidum*: a potent pharmacological macro fungus. *Current Pharmaceutical Biotechnology* 10, 717-742.
- Santos ARS, Calixto JB. (1997). Further evidence for the involvement of tachykinin receptor subtypes in formalin and capsaicin models of pain in mice. *Neuropeptides* 31, 381-389.
- Saraswat B, Visen PK, Agarwal DP. (2000). Ursolic acid isolated from *Eucalyptus tereticornis* protects against ethanol toxicity in isolated rat hepatocytes. *Phytotherapy Research* 14, 163-166.
- Scott DA, Wright CE, Angus JA. (2004). Evidence that CB-1 and CB-2 cannabinoid receptors mediate antinociception in neuropathic pain in the rat. *Pain* 109, 124-131.
- Sethi G, Ahn KS, Pandey MK, Aggarwal BB. (2007). Celastrol, a novel triterpene, potentiates TNFinduced apoptosis and suppresses invasion of tumor cells by inhibiting NF-kappaB-regulated gene products and TAK1-mediated NFkappaB activation. *Blood* 109, 2727-2735.
- Sethi G, Tergaonkar V. (2009). Potential pharmacological control of the NF-kappaB pathway. *Trends in Pharmacological Science* 30, 313-321.
- Shishodia S, Majumdar S, Banerjee S, Aggarwal BB. (2003). Ursolic acid inhibits nuclear factor kappaB activation induced by carcinogenic agents through suppression of I-kappa B-alpha kinase and p65 phosphorylation: correlation with down-regulation of cyclooxygenase 2, matrix metalloproteinase 9, and cyclin D1. *Cancer Research* 63, 4375-4383.
- Somova LI, Shode FO, Mipando M. (2004). Cardiotonic and antidysrhythmic effects of oleanolic and ursolic acids, methyl maslinate and uvaol. *Phytomedicine* 11, 121-129.
- Somova LO, Nadar A, Rammanan P, Shode FO. (2003). Cardiovascular, antihyperlipidemic and antioxidant effects of oleanolic and ursolic acids in experimental hypertension. *Phytomedicine* 10, 115-121.

- Subarnas A, Tadano T, Kisara K, Ohizumi O. (1993). An alpha-adrenoceptor-mediated mechanism of hypoactivity induced by beta-amyrin palmitate. *Journal of Pharmacy and Pharmacology* 45, 1006-1008.
- Sung B, Park B, Yadav VR, Aggarwal BB. (2010). Celastrol, a triterpene, enhances TRAIL-induced apoptosis through the down-regulation of cell survival proteins and up-regulation of death receptors. *The Journal of Biological Chemistry* 285, 11498-11507.
- Syrovets T, Buchele B, Krauss C, Laumonnier Y, Simmet T. (2005). Acetyl-boswellic acids inhibit lipopolysaccharide-mediated TNF-alpha induction in monocytes by direct interaction with IkappaB kinases. *Journal of Immunology* 174, 498-506.
- Tan SM, Li F, Rajendran P, Prem Kumar A, Hui KM, Sethi G. (2010). Identification of {beta}-escin as a novel inhibitor of signal transducer and activator of transcription 3/Janus-activated kinase 2 signaling pathway that suppresses proliferation and induces apoptosis in human hepatocellular carcinoma cells. *Journal of Pharmacol and Experimental Therapeutics* 334, 285-293.
- Tan Y, Yu R, Pezzuto JM. (2003). Betulinic acid-induced programmed cell death in human melanoma cells involves mitogen-activated protein kinase activation. *Clinical Cancer Research* 9, 2866-2875.
- Tang XL, Yang XY, Jung HJ, Kim SY, Jung SY, Choi DY, Park WC, Park H. (2009). Asiatic acid induces colon cancer cell growth inhibition and apoptosis through mitochondrial death cascade. *Biological and Pharmaceutical Bulletine* 32, 1399-1405.
- Taylor RF. (1984). Bacterial triterpenoids. Microbiological Reviews 48, 181-198.
- Tiedemann RE, Schmidt J, Keats JJ, Shi CX, Zhu YX, Palmer SE, Mao X, Schimmer AD, Stewart AK. (2009). Identification of a potent natural triterpenoid inhibitor of proteosome chymotrypsin-like activity and NF-kappaB with antimyeloma activity *in vitro* and *in vivo*. *Blood* 113, 4027-4037.
- Vitor CE, Figueiredo CP, Hara DB, Bento AF, Mazzuco TL, Calixto JB. (2009). Therapeutic action and underlying mechanisms of a combination of two pentacyclic triterpenes, alpha- and beta-amyrin, in a mouse model of colitis. *British Journal of Pharmacology* 157, 1034-1044.
- Vogel HG, De Souza ND. (1990). Effects of terpenoids isolated from *Centella asiatica* on granuloma tissue. *Acta Therapeutica* 16, 285-298.
- Wagner KH, Elmadfa I. (2003). Biological relevance of terpenoids: overview focusing on mono-, diand tetraterpenes. *Annals of Nutrition Metabolism* 47, e106.
- Won JH, Shin JS, Park HJ, Jung HJ, Koh DJ, Jo BG, Lee JY, Yun K, Lee KT. (2010). Anti-inflammatory effects of madecassic acid via the suppression of NF-kappaB pathway in LPS-induced RAW 264.7 macrophage cells. *Planta Medica* 76, 251-257.
- Wu CC, Chan ML, Chen WY, Tsai CY, Chang FR, Wu YC. (2005). Pristimerin induces caspasedependent apoptosis in MDA-MB-231 cells via direct effects on mitochondria. *Molecular Cancer Therapeutics* 4, 1277-1285.
- Wu X, Lee VC, Chevalier E, Hwang ST. (2009). Chemokine receptors as targets for cancer therapy. *Current Pharmaceutical Design* 15, 742-757.
- Xu R, Fazio GC, Matsuda SPT. (2004). On the origins of triterpenoid skeletal diversity. *Phytochemistry* 65, 261-291.
- Xu ZX, Liang J, Haridas V, Gaikwad A, Connolly FP, Mills GB, Gutterman JU. (2007). A plant triterpenoid, avicin D, induces autophagy by activation of AMP-activated protein kinase. *Cell Death Differentiation* 14, 1948-1957.
- Lia Y, Hea D, Zhanga X, Liua Z, Zhanga X, Donga L, et al., (2012). Protective effect of celastrol in rat cerebral ischemia model: Down-regulating p-JNK, p-c-Jun and NF-κB. *Brain Research* 1464, 8-13.
- Yildiz A, Gonul AS, Tamam L. (2002). Mechanism of actions of antidepressants: beyond the receptors. *Bulletin of Clinical Psychopharmacology* 12, 194-200.
- Yin MC. (2012). Anti-glycative potential of triterpenes: a mini-review. Biomedicine 2, 2-9.

- Yoo KY, Park SY. (2012). Terpenoids as potential anti-Alzheimer's disease therapeutics. *Molecules* 17, 3524-3538.
- Zhang C, Li B, Gaikwad AS, Haridas V, Xu Z, Gutterman, JU, Duvic M. (2008). Avicin D selectively induces apoptosis and downregulates p-STAT-3, bcl-2, and survivin in cutaneous T-cell lymphoma cells. *Journal of Investigative Dermatology* 128, 2728-2735.
- Zhang D, Xu L, Cao F, Wei T, Yang C, Uzan G, Peng B. (2010). Celastrol regulates multiple nuclear transcription factors belonging to HSP90's clients in a doseand cell type-dependent way. *Cell Stress Chaperones* 15, 939-946.
- Zhang L, Fu F, Zhang X, Zhu M, Wang T, Fan H. (2010). Escin attenuates cognitive deficits and hippocampal injury after transient global cerebral ischemia in mice via regulating certain inflammatory genes. *Neurochemistry International* 57, 119-127.
- Zhang P, Li H, Chen D, Ni J, Kang Y, Wang S. (2007). Oleanolic acid induces apoptosis in human leukemia cells through caspase activation and poly(ADPribose) polymerase cleavage. Acta Biochimica and Biophysica Sinica 39, 803-809.
- Zuco V, Supino R, Righetti SC, Cleris L, Marchesi E, Gambacorti-Passerini C, Formelli F. (2002). Selective cytotoxicity of betulinic acid on tumor cell lines, but not on normal cells. *Cancer Letters* 175, 17-25.